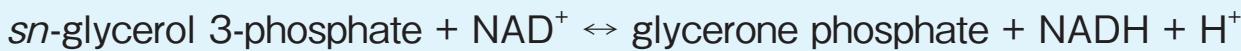


(Diagnostic Reagent Grade)

T-207

# GLYCEROL-3-PHOSPHATE DEHYDROGENASE [G3PDH II]

from *Saccharomyces cerevisiae*  
(*sn*-glycerol-3-phosphate:NAD<sup>+</sup> 2-oxidoreductase, EC 1.1.1.8)



## Preparation and Specification

Appearance : White lyophilized powder  
 Specific activity : More than 37.0 U/mg solid  
 Contaminants : NADH oxidase Less than 0.00015 %(U/U)

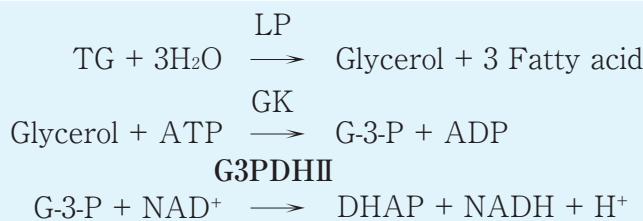
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## Properties

Substrate specificity	: The enzyme is specific for <i>sn</i> -glycerol 3-phosphate and NAD(H)
Molecular weight	: 45.6 kDa (SDS-PAGE)
Isoelectric point	: pH 4.8-5.3
Michaelis constant	: <i>sn</i> -glycerol 3-phosphate $1.4 \times 10^{-2}$ M (8.5 mM NAD) : NAD $4.6 \times 10^{-4}$ M (75 mM <i>sn</i> -glycerol 3-phosphate)
Optimum pH	: 9.0 Figure 1
pH stability	: 6.0-7.0 (37°C, 60 min) Figure 2
Optimum temperature	: 40°C Figure 3
Thermal stability	: Stable at 45°C and below (pH 6.5, 30 min) Figure 4
Inhibitor <sup>1)</sup>	: ATP, ADP, fructose 1,6-bisphosphate

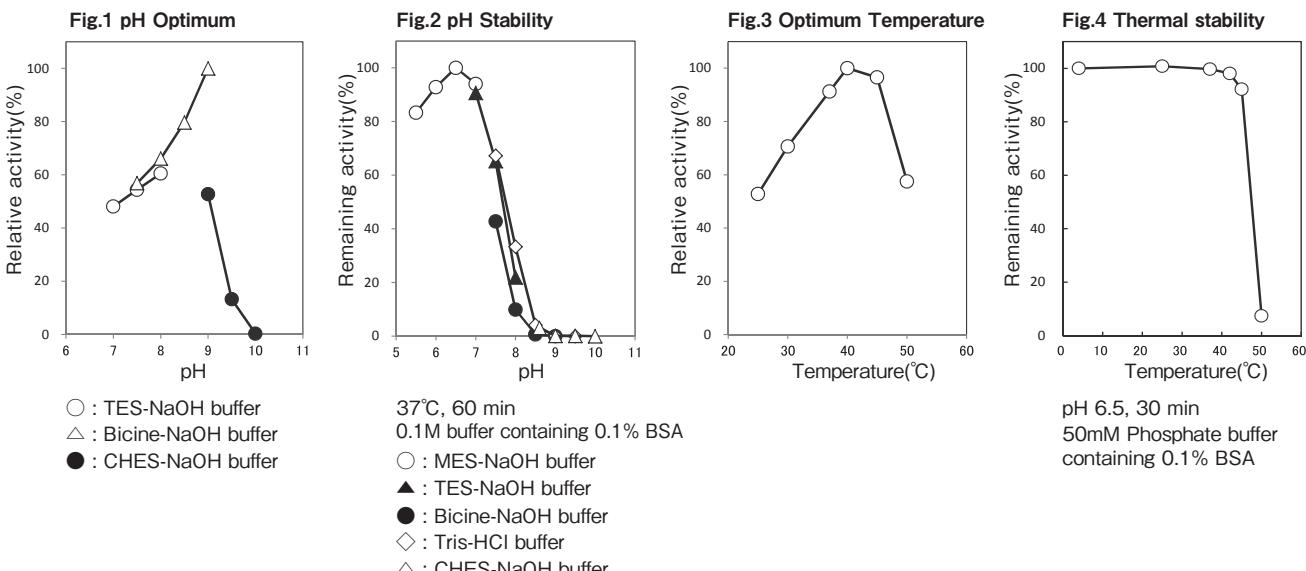
## Applications for Diagnostic Test

The enzyme is useful for enzymatic determination of triglyceride (TG) coupled with LP (Lipase; T-01, T-63, or T-116) and GK (Glycerol kinase; T-64 or T-223).



G-3-P: *sn*-Glycerol 3-phosphate

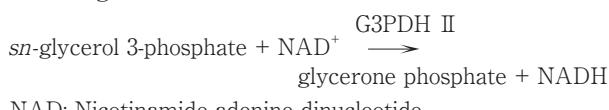
DHAP: Dihydroxyacetone phosphate



## Assay

### Principle

This assay is based on the increase in absorbance at 340 nm as the formation of NADH proceeds in the following reactions:



### Unit definition

One unit is defined as the amount of enzyme which generates 1 μmole of NADH per minute at 37 °C under the conditions specified in the assay procedure.

### Reagents

#### 1. Reaction mixture

Combine 1.632 g of Bicine with about 70 ml of distilled water, and adjust pH to about 8.5 at about 25 °C with 1N NaOH (Bicine will dissolve after adjusting pH). Add 4.727 g of Disodium Glycerophosphate 5.5-hydrate and 0.564 g (based on pure product) of NAD, dissolve them, add distilled water to make up to about 90 ml, and adjust pH to 8.7 at 25 °C with 1N NaOH. Finally add distilled water to make up to 100 ml.

#### 2. Enzyme dilution buffer

Dissolve 1.211 g of tris (hydroxymethyl) aminomethane with 800 ml of distilled water, and adjust pH to 7.5 at 25 °C with 1N HCl. Add 1 g of BSA, dissolve it, check the pH of the solution again, and add distilled water to make 1 L solution.

#### 3. Reagent

Bicine: Dojindo Laboratories #347-03282

Disodium Glycerophosphate 5.5-hydrate:

FUJIFILM Wako Pure Chemical Corporation  
Special grade #192-02055

Tris (hydroxymethyl) aminomethane:

Sigma Chemical Co. #T-1503

BSA: Millipore Fraction V pH 5.2 # 81-053

NAD: FUJIFILM Wako Pure Chemical Corporation  
#304-50443

### Enzyme solution

Accurately weigh about 20 mg of the sample and add enzyme dilution buffer to make a total of 20 ml. Dilute it with enzyme dilution buffer to adjust the concentration as required.

### Procedure

- Pipette accurately 2.00 ml of reaction mixture into a small test tube and preincubate at 37 °C.
- After 5 min. add accurately 20 μl of enzyme solution and mix to start the reaction at 37 °C.  
※ In the case of a test blank, add 20 μl of enzyme dilution buffer in place of enzyme solution.
- After starting the reaction, measure the rate of increase per minutes in absorbance at 340 nm. The rate must be measured within the linear portion of the absorbance curve.

$$\begin{aligned} \text{Absorbance sample : As/min} \\ \text{blank : Ab/min} \\ \Delta A/\text{min} = (\text{As}/\text{min} - \text{Ab}/\text{min}) \leq 0.15 \text{ Abs}/\text{min} \end{aligned}$$

### Calculation

$$\text{Activity (U/mg of powder)} = \frac{\Delta A/\text{min}}{6.22} \times \frac{2.02}{0.02} \times \frac{1}{X} \times D$$

6.22 : millimolar extinction coefficient of NADH at 340 nm  
(cm<sup>2</sup> / μmole)

2.02 : final volume (ml)

0.02 : volume of enzyme solution (ml)

X : concentration of the sample in enzyme solution (mg/ml)

D : times of dilution in enzyme solution

### Storage

Storage at -20 °C in the presence of a desiccant is recommended.

### References

- Albertyn J., van Tonder A., Prior BA. (1992) FEBS Lett., 308(2), 130-132. **72**, 972.
- Miura, M. et al. (1989) J. Clin. Lab. Inst. Reag., **12(5)**, 1005-1009.

# G3PDH II活性測定法

## I. 試薬液

### 1. 反応試薬混合液

Bicine 1.632 g を精製水約 70 ml に混合し、1N NaOH で約 pH8.5 (約 25℃) に調整する (pH 調整すると Bicine は溶解する)。さらにグリセロりん酸二ナトリウム 5.5 水和物 4.727 g と NAD 0.564 g を加えて溶解したあと、精製水を加えて全容を約 90 ml とし、1N NaOH で pH8.7 (25℃) に調整する (NAD は純度換算する)。最後に精製水で全容を 100 ml とする。

### 2. 酵素溶解希釈用液

トリス (ヒドロキシメチル) アミノメタン 1.211 g を精製水 800ml に溶解した後、1N HCl で pH7.5 (25℃) に調整し、BSA を 1 g 加えて溶解し、再度 pH を確認した後、精製水で 1 L とする。

### 3. 試薬

Bicine (ビシン) : 同仁化学製 #347-03282

グリセロりん酸二ナトリウム 5.5 水和物 :

富士フィルム和光純薬製 特級 #192-02055

トリス (ヒドロキシメチル) アミノメタン:

シグマ社製 #T-1503

BSA:Millipore 社製 Fraction V pH5.2 #81-053

NAD (ニコチンアミドアデニンジヌクレオチド・酸化型):

富士フィルム和光純薬製 #304-50443

## II. 酵素試料液

検品約 20 mg を精密に量り、酵素溶解希釈用液で溶解して全容 20 ml とする。

その液を酵素溶解希釈用液で適宜希釈する。

## III. 測定操作法

1. 小試験管に反応試薬混合液 2.00 ml を正確に分注し、37℃ で予備加温する。

2. 5分経過後、酵素試料液 20 μlを正確に加えて混和し、37℃ で反応を開始する。

※盲検は酵素試料液の代わりに酵素溶解希釈用液 20 μl を加える。

3. 反応開始後、340 nmにおける吸光度を測定して直線的に反応している1分間当たりの吸光度変化を求める。求められた吸光度変化を試料液についてはAs/min、盲検液についてはAb/min とする。

$$\Delta A/min = (As/min - Ab/min) \leq 0.15 Abs/min$$

## IV. 計算

$$\text{活性 (U/mg)} = \frac{\Delta A/min}{6.22} \times \frac{2.02}{0.02} \times \frac{1}{X} \times D$$

6.22 : NADH の 340nm におけるミリモル分子

吸光係数 (cm<sup>2</sup>/ μmol)

2.02 : 反応総液量 (ml)

0.02 : 反応に供した酵素試料液量 (ml)

X : 酵素試料液中の検品濃度 (mg/ml)

D : 酵素試料液の希釈倍率