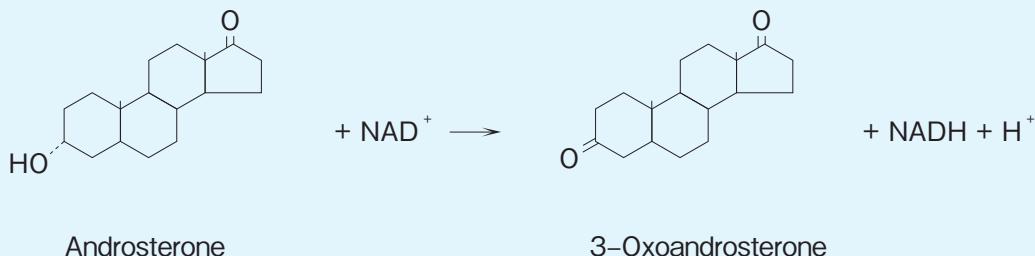


(Diagnostic Reagent Grade)

T-58

3α -HYDROXYSTEROID DEHYDROGENASE [3α -HSD II]

from *Pseudomonas* sp. $(3\alpha$ -Hydroxysteroid: NAD $^+$ oxidoreductase, EC 1.1.1.50)

Preparation and Specification

Appearance : White amorphous powder, lyophilized
Specific activity : More than 30 U/mg solid

Properties

Substrate specificity	: See Table 1	
Molecular weight	: 41 kDa (gel filtration)	
Isoelectric point	: pH 4.8 \pm 0.2	
Michaelis constant	: Androsterone 2.1×10^{-4} M	
Optimum pH	: 8.0-10.0	Figure 1
pH stability	: 6.0-10.0 (37°C, 10 min)	Figure 2
Optimum temperature	: 50°C (Phosphate buffer)	Figure 3
Thermal stability	: Stable at 50°C and below (pH 8.0, 10 min)	Figure 4
Effect of metal ions	: See Table 2	
Effect of detergents	: See Table 3	
Inhibitor	: MnCl ₂	

Applications for Diagnostic Test

This enzyme is useful for enzymatic cycling determination of **bile acid** when coupled with thio-NAD and NADH.

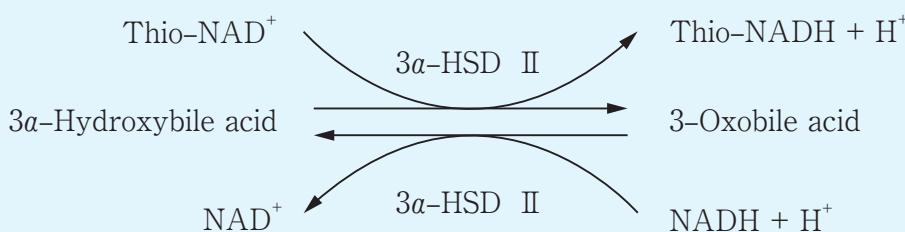


Table 1. Substrate specificity

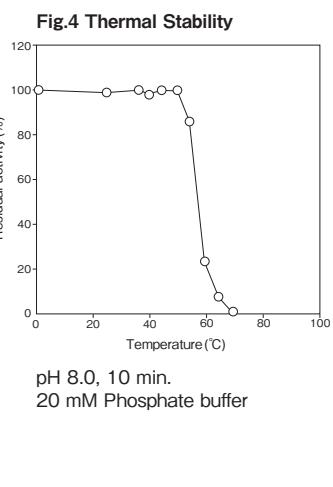
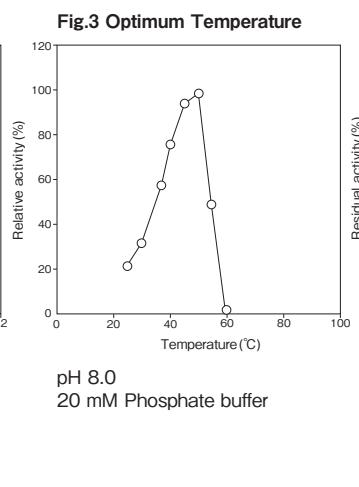
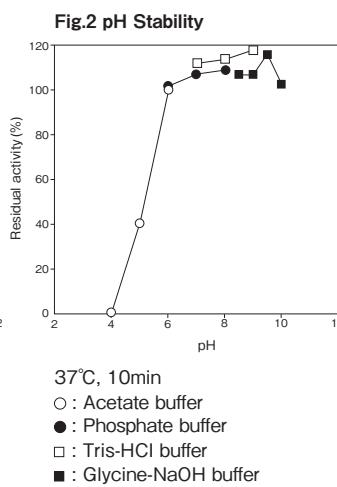
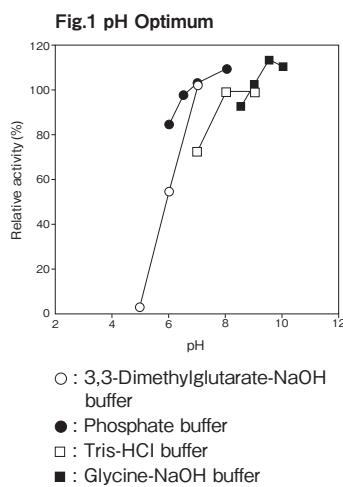
Substrate (1mM)	Relative activity (%)
Cholic acid	100
Androsterone	131
Deoxycholic acid	115
Chenodeoxycholic acid	89.0
Glycocholic acid	103
Taurocholic acid	99.0
Taurodeoxycholic acid	128

Table 2. Effect of metal ions on 3 α -HSD II activity

Metal ion (10mM)	Relative activity (%)
None	100
NaCl	105
KCl	102
LiCl	101
MgCl ₂	106
MnCl ₂	16.0
CaCl ₂	105

Table 3. Effect of detergents on 3 α -HSD II activity

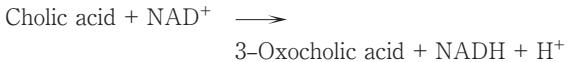
Detergent (0.1%)	Relative activity (%)
None	100
Triton X-100	71.0
Triton X-305	71.0
Triton X-114	71.0
Adekanol SO-120	104
Adekanol NP-720	75.0
Adekanol B-795	78.0
Emulgen B-66	75.0
Emulgen 911	76.0
Emulgen 709	84.0
Emulgen 810	50.0
Emulgen 109P	112
Rheodol 460	71.0
Rheodol TWL-103	72.0



Assay

Principle

The assay is based on the increase in absorbance at 550 nm as formazan dye is formed in the following reactions:



DI



NTB: Nicotinamide adenine dinucleotide,

NTB: Nitrotetrazolium blue

DI: Diaphorase

Unit definition

One unit is defined as the amount of enzyme which oxidizes 1 μ mole of cholic acid to 3-oxocholic acid per minute at 37°C under the conditions specified in the assay procedure.

Reagents

1. Reaction mixture

10 mM NAD solution 0.05 ml

0.25% (W/V) NTB solution 0.05 ml

100 U/ml DI solution 0.025 ml

2% (W/V) Triton X-100 solution 0.10 ml

0.2 M Tris-HCl buffer pH 8.0 0.10 ml

Distilled water 0.175 ml

1): 100 U/ml DI solution

Dissolve 100 U of DI with 1 ml of 10 mM Tris-HCl buffer pH 8.0.

2. Substrate solution (20 mM Androsterone)

Dissolve 23 mg of androsterone with 4 ml of methanol.

3. Reaction stopper

0.5% (W/V) Sodiumdodecyl sulfate (SDS) solution

4. Enzyme dilution buffer

10 mM Tris-HCl buffer pH 8.0

5. Reagents

NAD: NACALAI TESQUE, INC. #24334-84

NTB: Dojindo Laboratories #344-02033
 DI: Nagase Diagnostics Co., Ltd. #T-06
 Triton X-100: The Dow Chemical Company
 Androsterone: Sigma Chemical Co. #A-9755
 SDS (Sodium Dodecyl Sulfate):
 NACALAI TESQUE, INC. Extra pure #31606-75

■ Enzyme solution

Accurately weigh about 20 mg of the sample and add enzyme dilution buffer to make a total of 20 ml. Dilute it with enzyme dilution buffer to adjust the concentration as required.

■ Procedure

- Pipette accurately 0.5 ml of reaction mixture into a small test tube, then add 20 μ l of enzyme solution into the same test tube and preincubate at 37°C.
※ In the case of a test blank, add 20 μ l of enzyme dilution buffer in place of enzyme solution.
- After 5 min, add exactly 25 μ l of substrate solution and mix to start the reaction at 37°C.
- At 5 min after starting the reaction, add 2.5 ml of the reaction stopper to stop the reaction.
- Measure the absorbance at 550 nm.

$$\begin{array}{ll} \text{Absorbance} & \text{sample : As} \\ & \text{blank : Ab} \\ \Delta A = (As - Ab) & \leq 0.20 \text{ Abs} \end{array}$$

■ Calculation

$$\text{Activity (U/mg of powder)} = \frac{\Delta A / 5}{16.7} \times \frac{3.045}{0.02} \times \frac{1}{X}$$

3 α -HSD II 活性測定法 (Japanese)

I. 試薬液

1. 反応試薬混合液

10mM NAD 溶液	0.05 ml
0.25% (W/V) NTB 溶液	0.05 ml
100U/ml DI 溶液 ¹⁾	0.025 ml
2% (W/V) トリトン X-100 溶液	0.10 ml
0.2M トリス-HCl 緩衝液 pH8.0	0.10 ml
精製水	0.175 ml
1):100U/ml DI 溶液 DI 100 単位 (U) を 10mM トリス-HCl 緩衝液 pH8.0 1ml で溶解する。	

2. 基質溶液 (20mM アンドロステロン溶液)

アンドロステロン 23mg を MeOH4ml で溶解する。

3. 反応停止液

0.5% (W/V) SDS 溶液

4. 酵素溶解希釈用液

10mM トリス-HCl 緩衝液 pH8.0

5. 試薬

NAD (ニコチンアミドアデニジヌクレオチド):
ナカライトスク製 #24334-84

NTB (ニトロテトラゾリウムブルー):
同仁化学製 #344-02033

DI (ジアフォラーゼ): ナガセダイアグノスティックス製 #T-06
トリトン X-100:Dow Chemical 製

アンドロステロン:シグマ製 #A-9755

SDS (ドデシル硫酸ナトリウム):
ナカライトスク製 一級 #31606-75

16.7 : millimolar extinction coefficient of NTBH₂ at 550 nm
(cm²/ μ mole)
5 : reaction time (min)
3.045 : final volume (ml)
0.02 : volume of enzyme solution (ml)
X : concentration of the sample in enzyme solution
(mg/ml)

Storage

Storage at -20°C in the presence of a desiccant is recommended.

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II. 酵素試料液

検品約 20mg を精密に量り、酵素溶解希釈用液で溶解して全容 20ml とする。

その液を酵素溶解希釈用液で適宜希釈する。

III. 測定操作法

1. 小試験管に反応試薬混合液 0.5ml を正確に分注し、後に酵素試料液 20 μ l を正確に分注して 37°C で予備加温する。

※盲検は酵素試料液の代わりに酵素溶解希釈用液 20 μ l を加える。

2. 5 分経過後、基質溶液 25 μ l を正確に加えて混和し、37°C で反応を開始する。

3. 5 分経過後、反応停止液 2.50ml を正確に加えて混和し、反応を停止する。

4. 550nm における吸光度を測定する。
求められた吸光度を試料液は As、盲検液は Ab とする。

$$\Delta A = (As - Ab) \leq 0.20 \text{ Abs}$$

IV. 計算

$$\text{活性 (U/mg)} = \frac{\Delta A / 5}{16.7} \times \frac{3.045}{0.02} \times \frac{1}{X}$$

16.7 : NTBH₂ の 550nm におけるミリモル分子吸光係数
(cm²/ μ mole)

5 : 反応時間 (min)

3.045 : 反応総液量 (ml)

0.02 : 反応に供した酵素試料液量 (ml)

X : 酵素試料液中の検品濃度 (mg/ml)