

(Diagnostic Reagent Grade)

T-16

A

# ACYL-CoA SYNTHETASE [ACS]

from *Pseudomonas fragi*  
 (Acid: CoA ligase (AMP forming), EC 6.2.1.3)



## Preparation and Specification

Appearance : White amorphous powder, lyophilized  
 Specific activity : More than 2 U/mg solid

## Properties

Substrate specificity	: See Table 1	
Molecular weight	: 60 kDa (Sephadex G-150) , 62 kDa (SDS-PAGE)	
Isoelectric point	: pH 5.2	
Michaelis constants	: Palmitic acid $1.1 \times 10^{-5}\text{M}$ ATP $1.7 \times 10^{-4}\text{M}$ CoA $3.2 \times 10^{-4}\text{M}$	
Optimum pH	: Palmitic acid      7.7	Figure 1
	: Serum fatty acids      7.7	Figure 2
pH stability	: 6.0-8.0 (37°C, 2 hr)	Figure 3
Thermal stability	: Stable at 50°C and below : (pH 7.5, 10 min)	Figure 4
Storage stability	: At least one year at -20°C	Figure 5
Effect of various chemicals	: See Table 2 and Table 3	
Stabilizer	: ATP	
Activator	: Triton X-100	

## Applications for Diagnostic Test

This enzyme is useful for enzymatic determination of **fatty acid** when coupled with Acyl-CoA oxidase (T-17) .

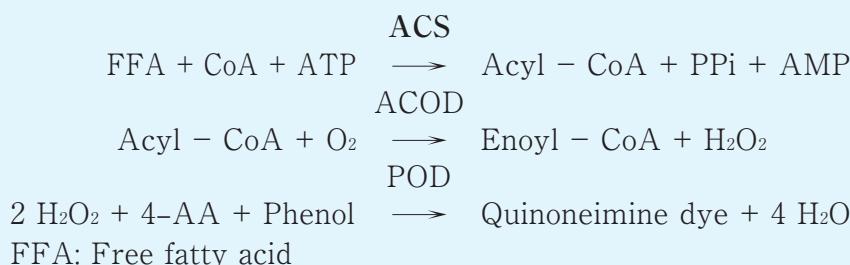


Table 1. Substrate specificity (Fatty acids)

Substrate		Relative activity (%)	Km value ( $10^{-5}$ M)
Caproic acid	(6:0)	38	
Caprylic acid	(8:0)	64	3.7
Capric acid	(10:0)	24	0.5
Lauric acid	(12:0)	21	0.95
Myristic acid	(14:0)	40	0.71
Palmitic acid	(16:0)	66	1.10
Stearic acid	(18:0)	78	3.0
Arachidic acid	(20:0)	100	
Myristoleic acid	(14:1)	35	
Palmitoleic acid	(16:1)	40	
Palmitelaidic acid	(16:1)	48	
Oleic acid	(18:1)	78	0.91
Elaidic acid	(18:1)	60	
Linoleic acid	(18:2)	57	0.34
Linolenic acid	(18:3)	62	1.10
Arachidonic acid	(20:4)	63	
Erucic acid	(22:1)	92	
Nervonic acid	(24:1)	9	

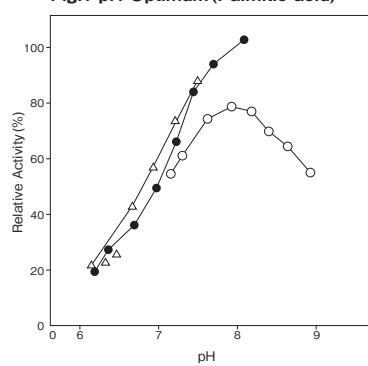
Table 2. Effect of detergents on ACS activity

Detergent (%)	Relative activity (%)
None	100
Deoxycholate	69.6
	41.1
SDS	0
Cetyltrimethylammoniumchloride	96.4
	0
Cetylpyridinium chloride	96.4
	0
Sarcosinate PN	64.3
	0

Table 3. Effect of metal ions on ACS activity

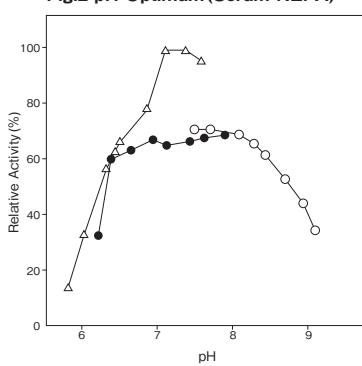
Metal ion	Concentration	Relative activity (%)
None	-	38
KCl	0.1M	29
NaCl	0.1M	35
LiCl	0.1M	26
NH <sub>4</sub> Cl	0.1M	29
MgCl <sub>2</sub>	1mM	100
CaCl <sub>2</sub>	1mM	94
ZnCl <sub>2</sub>	1mM	31
BaCl <sub>2</sub>	1mM	38
MnCl <sub>2</sub>	1mM	117
CuCl <sub>2</sub>	1mM	0
NiCl <sub>2</sub>	1mM	87
EDTA	1mM	0
1mM MgCl <sub>2</sub> + CaCl <sub>2</sub>	1mM	103
1mM MgCl <sub>2</sub> + ZnCl <sub>2</sub>	1mM	55
1mM MgCl <sub>2</sub> + CuCl <sub>2</sub>	1mM	0
1mM MgCl <sub>2</sub> + NiCl <sub>2</sub>	1mM	99
1mM MgCl <sub>2</sub> + BaCl <sub>2</sub>	1mM	100
1mM MgCl <sub>2</sub> + MnCl <sub>2</sub>	1mM	117

Fig.1 pH Optimum (Palmitic acid)



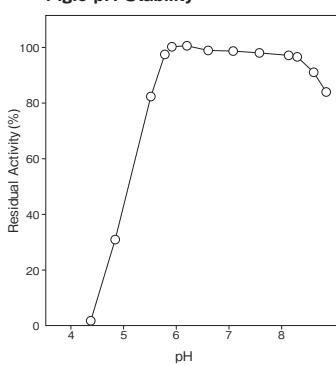
△ : 3,3-Dimethylglutarate-NaOH buffer  
● : Phosphate buffer  
○ : Tris-HCl buffer

Fig.2 pH Optimum (Serum NEFA)

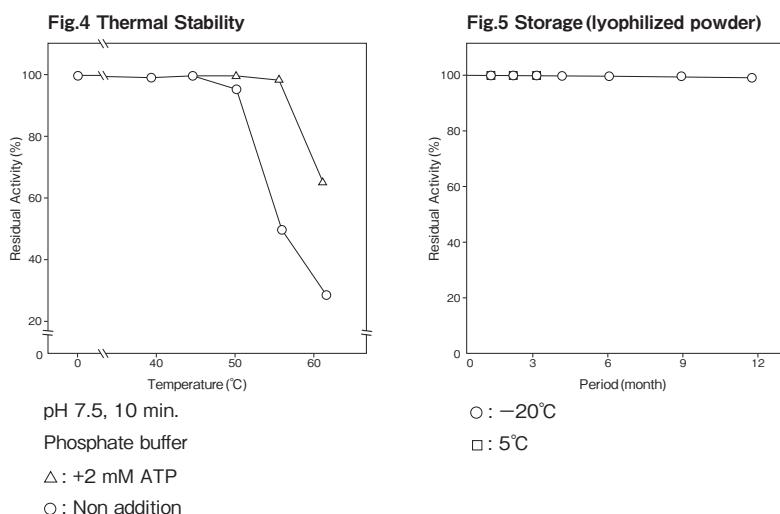


△ : 3,3-Dimethylglutarate-NaOH buffer  
● : Phosphate buffer  
○ : Tris-HCl buffer

Fig.3 pH Stability



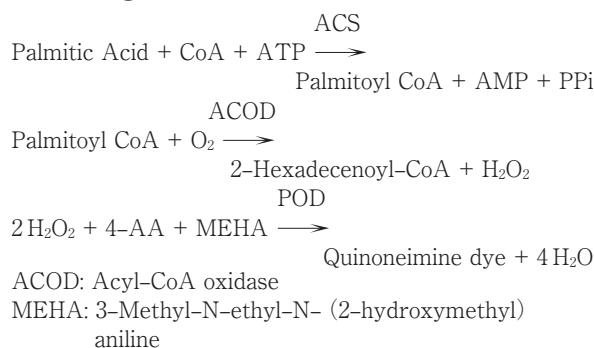
37°C, 2 hr  
pH 4.0–6.5 3,3-Dimethylglutarate-NaOH buffer  
pH 6.5–7.5 Phosphate buffer  
pH 7.5–9.0 Tris-HCl buffer



## Assay

### ■ Principle

The assay is based on the increase in absorbance at 550 nm as the formation of quinoneimine dye proceeds in the following reactions:



### ■ Unit definition

One unit is defined as the amount of enzyme which converts 1 μmole of fatty acid to acyl-CoA per minute at 37°C under the conditions specified in the assay procedure.

### ■ Reagents

#### 1. Reaction mixture for the first reaction

0.2 M KH <sub>2</sub> PO <sub>4</sub> –K <sub>2</sub> HPO <sub>4</sub> buffer pH 7.5	0.20 ml
10 mM ATP solution pH 7.5	0.10 ml
10 mM MgCl <sub>2</sub> solution	0.10 ml
1 mM Palmitic acid-5 % (W/V) Triton X-100 solution pH 7.5 <sup>1)</sup>	0.20 ml
Distilled water	0.35 ml
10 mM CoA solution pH 6.5 <sup>2)</sup>	0.05 ml

1): 1 mM Palmitic acid-5 % (W/V) Triton X-100 solution pH 7.5

Dissolve 26 mg of palmitic acid with 90 ml of 5% (W/V) Triton X-100, adjust pH to 7.5 at 25°C with 4 N NaOH, add 5% (W/V) Triton X-100 to make a total of 100 ml.

2): 10 mM CoA solution pH 6.5

Dissolve 154 mg (purity calculation) of CoA with 15 ml of distilled water, adjust pH to 6.5 at 25°C with 4 N NaOH, and add distilled water to make a total of 20 ml.

#### 2. Reaction mixture for the second reaction

0.2 M KH <sub>2</sub> PO <sub>4</sub> –K <sub>2</sub> HPO <sub>4</sub> buffer pH 7.5	0.50 ml
20 mM NEM	0.10 ml
15 mM 4-AA solution	0.30 ml
0.3% (W/V) MEHA solution pH 5.8	0.25 ml
100 U/ml POD solution <sup>3)</sup>	0.10 ml
0.5% (W/V) NaN <sub>3</sub> solution	0.10 ml
Distilled water	0.55 ml
120 U/ml ACOD solution <sup>4)</sup>	0.10 ml

NEM: N-Ethylmaleimide

3): 100 U/ml POD solution

Dissolve 1,000 U (PPU) of POD with 10 ml of distilled water.

4): 120 U/ml ACOD solution

Dissolve 1,200 U of ACOD with 10 ml of ACOD dilution buffer<sup>\*\*</sup>.

\*\*): ACOD dilution buffer

Dissolve 1.36 g of KH<sub>2</sub>PO<sub>4</sub> and 1.82 g of ATP with distilled water, adjust pH to 7.0 with 4 N NaOH, add 10 ml of 1 mM FAD, and finally add distilled water to make a total of 1 L.

#### 3. Enzyme dilution buffer

10 mM KH<sub>2</sub>PO<sub>4</sub>–K<sub>2</sub>HPO<sub>4</sub> buffer(pH 7.5) containing 2 mM ATP, 0.5% (W/V) BSA and 0.1% (W/V) Triton X-100.

#### 4. Reagents:

ATP (2Na · 3H<sub>2</sub>O): Kyowa Hakko Co., Ltd.

CoA (Coenzyme A): KOHJIN

Palmitic acid :

FUJIFILM Wako Pure Chemical Corporation #169-00105

Triton X-100: The Dow Chemical company

NEM: FUJIFILM Wako Pure Chemical Corporation

Special grade #058-02061

4-AA: NACALAI TESQUE, INC.

Special grade #01907-52

MEHA: Tokyo Kasai Kogyo Co., Ltd. #E0220

POD: Sigma Chemical Co. Type II #P-8250

ACOD: Nagase Diagnostics Co., Ltd. #T-17

FAD (2Na): Kyowa Hakko Co., Ltd.

BSA: Millipore Fraction V pH 5.2 #81-053

ATP: Adenosine triphosphate  
FAD: Flavine adenine dinucleotide

### ■ Enzyme solution

Weigh about 20 mg of test sample exactly and add enzyme dilution buffer to make a total of 20 ml. Dissolve it with enzyme dilution buffer to adjust the concentration to within 0.04–0.06 U/ml.

### ■ Procedure

- Pipette accurately 1.0 ml of reagent mixture for the first reaction into a small test tube and preincubate at 37°C.
- After 5 min, add exactly 50  $\mu$ l of enzyme solution and mix to start the first reaction at 37°C.  
※ In the case of a test blank, add 50  $\mu$ l of enzyme dilution buffer in place of enzyme solution.
- After 10 min, add 2.0 ml of reagent mixture for the second reaction to stop the first reaction and mix to start the second reaction at 37°C.
- After 5 min, measure the absorbance at 550 nm.

Absorbance sample : As

blank : Ab

$$\Delta A = (As - Ab) \leq 0.30 \text{ Abs}$$

### ■ Calculation

$$\text{Activity (U/mg)} = \frac{\Delta A / 10}{32.0 \times 1/2} \times \frac{3.05}{0.05} \times \frac{1}{X}$$

## ACS 活性測定法 (Japanese)

### I. 試薬液

#### 1. 第一反応試薬混合液

0.2M KH <sub>2</sub> PO <sub>4</sub> -K <sub>2</sub> HPO <sub>4</sub> 緩衝液 pH7.5	0.20 ml
10mM ATP 溶液 pH7.5	0.10 ml
10mM 塩化マグネシウム溶液	0.10 ml
1mM パルミチン酸-5% (W/V) トリトン X-100 溶液 pH7.5 <sup>1)</sup>	0.20 ml
精製水	0.35 ml
10mM CoA 溶液 pH6.5 <sup>2)</sup>	0.05 ml
1): 1mM パルミチン酸-5% (W/V) トリトン X-100 溶液 pH7.5 パルミチン酸 26mg を 5% (W/V) トリトン X-100 溶液 90ml で加温溶解した後、4N NaOH で pH7.5 (25°C) に調整し、5% (W/V) トリトン X-100 溶液で全容 100ml とする。	
2): 10mM CoA 溶液 pH6.5 CoA 154mg (純度換算) を精製水 15ml に溶解し た後、4N NaOH で pH6.5 (25°C) に調整し、精製 水で全容 20ml とする。	

#### 2. 第二反応試薬混合液

0.2M KH <sub>2</sub> PO <sub>4</sub> -K <sub>2</sub> HPO <sub>4</sub> 緩衝液 pH7.5	0.50 ml
20mM NEM 溶液	0.10 ml
15mM 4-AA 溶液	0.30 ml
0.3% (W/V) MEHA 溶液 pH5.8	0.25 ml
100U/ml POD 溶液 <sup>3)</sup>	0.10 ml

32.0 : millimolar extinction coefficient of quinoneimine dye at 550 nm ( $\text{cm}^2 / \mu\text{mole}$ )

1/2 : a multiplier derived from the fact that 2 mole of H<sub>2</sub>O<sub>2</sub> produces 1 mole of quinoneimine dye

10 : reaction time (min)

3.05 : final volume (ml)

0.05 : volume of enzyme solution (ml)

X : concentration of ACS in enzyme solution (mg/ml)

### Storage

Storage at -20°C in the presence of a desiccant is recommended. Enzyme activity will be retained for at least one year under this condition.

### References

- Yamada, H., Shimizu, S. and Tani, Y. (1980) Vitamin (Japanese), **54**, 489.
- Shimizu, S., Inoue, K., Tani, Y. and Yamada, H. (1980) Anal. Biochem., **107**, 193.
- Okabe, H., Uji, Y., Nagashima, K. and Noma, A. (1980) Clin. Chem., **26**, 1540.
- Shimizu, S., Inoue, K., Tani, Y. and Yamada, H. (1979) Anal. Biochem., **98**, 341.

0.5% (W/V) NaN<sub>3</sub> 溶液

0.10 ml

精製水

0.55 ml

120U/ml ACOD 溶液<sup>4)</sup>

0.10 ml

3): 100U/ml POD 溶液

POD 1,000 単位(PPU) を精製水 10ml で溶解する。

4): 120U/ml ACOD 溶液

ACOD 1,200 単位 (U) を ACOD 希釀緩衝液<sup>5)</sup> 10ml で溶解する。

※): ACOD 希釀緩衝液

KH<sub>2</sub>PO<sub>4</sub> 1.36g と ATP 1.82g を精製水に溶解した後、4N NaOH で pH7.0 (25°C) に調製し、さらに 1mM FAD 溶液 10ml を加えて精製水で全容 1L とする。

#### 3. 酵素溶解希釀用液

2mM ATP と 0.5% (W/V) BSA 及び 0.1% (W/V) トリトン X-100 を含む 10mM KH<sub>2</sub>PO<sub>4</sub>-K<sub>2</sub>HPO<sub>4</sub> 緩衝液 pH7.5

#### 4. 試薬

ATP (アデノシン三リン酸・2Na·3H<sub>2</sub>O):

協和発酵製

CoA (コエンザイム A): 興人製

パルミチン酸: 富士フイルム和光純薬製 特級

#169-00105

トリトン (トリトン X-100): Dow Chemical 製

NEM(N-エチルマレイミド):

富士フイルム和光純薬製 特級 #058-02061

4-AA: ナカライトスク製 特級 #01907-52

MEHA [N-エチル-N-2-ヒドロキシエチル-m-トルイジン]:

東京化成製 #E0220

POD:シグマ製 Type II #P-8250

ACOD (アシル-CoA 酸化酵素):

ナガセダイアグノスティックス製 #T-17

FAD (フラビンアデニンジヌクレオチド・2Na):

協和発酵製

BSA: Millipore 製 Fraction V pH5.2 #81-053

## II. 酵素試料液

検品約 20mg を精密に量り、酵素溶解希釈用液に溶解して全容 20ml とする。

その液を酵素溶解希釈用液で 0.04~0.06U/ml 濃度となるように適宜希釈する。

## III. 測定操作法

- 小試験管に第一反応試薬混合液 1.0ml を正確に分注して 37°C で予備加温する。

- 5 分経過後、酵素試料液 50 μl を加えて混和し、37°C で反応を開始する。

※盲検は酵素試料液の代わりに酵素溶解希釈用液 50 μl を加える。

- 10 分経過後、第二反応試薬混合液 2.0ml を正確に加えて混和し、第一反応を停止させ、37°C で第二反応を開始する。

- 5 分経過後、550nm における吸光度を測定する。求められた吸光度を試料液は As、盲検液は Ab とする。

$$\Delta A = (As - Ab) \leq 0.30 \text{ Abs}$$

## IV. 計算

$$\text{活性 (U/mg)} = \frac{\Delta A / 10}{32.0 \times 1/2} \times \frac{3.05}{0.05} \times \frac{1}{X}$$

32.0: キノンイミン色素の 550nm におけるミリモル分子吸光係数 (cm<sup>2</sup> / μmole)

1/2: H<sub>2</sub>O<sub>2</sub> 2 モルからキノンイミン色素 1 モルが生成することによる係数

10 : 反応時間 (min)

3.05 : 反応総液量 (ml)

0.05 : 反応に供した酵素試料液量 (ml)

X : 酵素試料液中の検品濃度 (mg/ml)